



BRITISH MEDICAL JOURNAL



British Medical Journal Volume-1, No 2
10.5281/zenodo.5221612

British Medical Journal

Volume 1, No 2., 2021

Internet address: <http://ejournals.id/index.php/bmj>

E-mail: info@ejournals.id

Published by British Medical Journal

Issued Bimonthly

3 knoll drive. London. N14 5LU United Kingdom

+44 7542 987055

Chief Editor

Dr. Fiona Egea

Requirements for the authors.

The manuscript authors must provide reliable results of the work done, as well as an objective judgment on the significance of the study. The data underlying the work should be presented accurately, without errors. The work should contain enough details and bibliographic references for possible reproduction. False or knowingly erroneous statements are perceived as unethical behavior and unacceptable.

Authors should make sure that the original work is submitted and, if other authors' works or claims are used, provide appropriate bibliographic references or citations. Plagiarism can exist in many forms - from representing someone else's work as copyright to copying or paraphrasing significant parts of another's work without attribution, as well as claiming one's rights to the results of another's research. Plagiarism in all forms constitutes unethical acts and is unacceptable. Responsibility for plagiarism is entirely on the shoulders of the authors. Significant errors in published works. If the author detects significant errors or inaccuracies in the publication, the author must inform the editor of the journal or the publisher about this and interact with them in order to remove the publication as soon as possible or correct errors. If the editor or publisher has received information from a third party that the publication contains significant errors, the author must withdraw the work or correct the errors as soon as possible.

OPEN ACCESS

Copyright © 2021 by British Medical Journal

British Medical Journal Volume-1, No 2

THE ROLE OF GENE POLYMORPHISM IN THE DEVELOPMENT OF ACUTE PANCREATITIS

Sabirova R.A.¹, Shukurov I.B.²

¹Tashkent Medical Academy

²Bukhara State Medical Institute

Abstract. The article analyzes scientific works on the role of gene polymorphism in the development of acute pancreatitis. The etiopathogenetic factors of destructive forms of acute pancreatitis are being studied for the purpose of predicting it and early surgical treatment. The authors emphasize the analysis of factors influencing the development of chronic pancreatitis.

Keywords. Pancreatitis, mutation, severe form, pancreatonecrosis, genetic status.

Introduction. In Russia, acute pancreatitis is one of the most common acute surgical diseases [1, 335 p.]. In different regions of the country, the number of patients hospitalized with a similar diagnosis varies from 38 to 95 people per 100,000 population and continues to grow every year [2, 55-59, 4-14]. It should be noted that in 80% of patients, acute pancreatitis occurs easily and spontaneously resolves within a week [3, P. 93–101.]. In other cases, severe forms of the disease develop (destructive, necrotizing pancreatitis), the mortality rate in which, according to various data, varies from 7 to 50 %, depending on the severity and prevalence of the process, averaging 20-30 %. [4, P. 565–573]. With infected pancreatic necrosis, which occurs in 40-70 % of cases, with a severe course of the disease, the mortality rate reaches 85 %, and with fulminant pancreatitis up to 100 % [5, p. 373-379.]. e

The increase in the incidence of pancreatitis is due, on the one hand, the characteristics of the diet, increasing abuse of alcohol and its surrogates, the prevalence of gallstone disease and, as a consequence, the increase in the absolute number of patients, and on the other hand, improvement in clinical, laboratory and instrumental diagnosis of the disease [6, p. 247-250].

In this regard, it is of interest to study the etiopathogenetic factors of destructive forms of acute pancreatitis in order to predict it and to treat it early. A review of the data of the modern literature indicates that researchers pay close attention to the genetic risk factors for pancreatitis.

The first ideas of genetic determination of predisposition to pancreatitis were expressed in the middle of the twentieth century. [7, P. 247-250.]. The disease is characterized by periodic attacks of acute pancreatitis in the absence of known provoking factors, debuts in childhood and is found in at least two other family members. Thus, L. Le Bodic et al. [8, P. 1504-1510.], having studied 249 family members from eight generations, found that this nosology is inherited by an autosomal dominant type with incomplete (80 %) penetrance. These works marked the beginning of the study of genetic changes associated with pancreatitis.

In recent years, researchers have been paying attention to the genetic predisposition to severe forms of the disease [9, p. 20-25.]. In this case, molecular genetic research methods are used to identify groups of patients who are determined

to develop severe forms of acute pancreatitis with septic complications. Many studies are devoted to the identification of gene polymorphism in chronic and acute pancreatitis. When analyzing the factors affecting the development of chronic pancreatitis, mutations in the cystic fibrosis gene (CFTR-transmembrane regulator of cystic fibrosis conduction), a pancreatic secretory trypsin inhibitor, were revealed. The literature also describes other mutations in genes that affect the state of the pancreas-SPINK1 (serine protease inhibitor kazal type 1), genes responsible for the synthesis of alcohol dehydrogenase and alpha-1 – antitrypsin. In the development of severe forms of pancreatitis, the authors note the role of polyorphism in the genes of tumor necrosis factor-2 and TNF-alpha (tumor necrosis factor) and IL-8 in [10, P. 247-250.]. A method for determining the severity of acute idiopathic pancreatitis based on the detection of heterozygous mutations in the SPINK1, PRSS1 and CFTR genes is described [11, p. 169-177.].

One of the most studied here is the cationic trypsinogen gene PRSS1 – over the past 15 years, many mutations associated with both hereditary and idiopathic and alcoholic pancreatitis have been described (for example, R122H, N21I, R116C, N29T, R122C, E79K, etc.) [12, P. 247–250].

Interaction between trypsinogen isoforms in genetically determined pancreatitis: mutation E79K in cationic trypsin (PRSS1) causes increased transactivation of anionic trypsinogen (PRSS2) // *Hum. Mutat.* 2004. Vol. 23, No. 1. P. 22-31.]. All of them lead to an acceleration of the conversion of trypsinogen to trypsin, which triggers a cascade of enzymatic reactions and serves as the pathogenetic basis for acute recurrent pancreatitis [13, P. 8–15.].

Another equally important protein that regulates trypsin is a serine protease inhibitor of the Kazal-1 type (SPINK1). Since the protection of the pancreas is provided by the balance between trypsin and its inhibitor, pancreatitis can develop not only with excessive activation of trypsinogen, but also with insufficient trypsin-binding ability of the inhibitor. In the case of damage to the SPINK1 gene, the inhibitory activity of the protein decreases, which leads to a violation of the inactivation of trypsin in the pancreatic tissue. Excess trypsin triggers the activation of other pancreatic enzymes with proteolytic necrosis of the gland tissue [4, P. 775–778.].

Mutations in the SPINK1 gene are found in 20-23% of patients with hereditary pancreatitis, which is several times higher than the frequency of the disease in the general population [15, P. 324–329.]. In the study of Yu. A. Kucheryavy, SPINK1 mutations were detected in all forms of chronic pancreatitis, except for autoimmune. The most common mutations of this gene associated with idiopathic chronic pancreatitis are N34S (replacement of asparagine with serine in codon 34) and P55S [16, P. 675-681.]. Mutations of the serine protease inhibitor, Kazal type 1 gene, in patients with idiopathic chronic pancreatitis // *Am. J. Gastroenterol.* 2002. Vol. 97, No. 5. P. 1133-1137.]. It is also shown that such genetic damage is a factor predisposing to alcoholic pancreatitis [17, P. 687–692]. Other mutations of the SPINK1 gene were also detected: R67C, R65Q, Y1092X, M1T, intron mutations c. 27deIC and C. 871G>A. The phenotypic manifestations of these genetic changes

have not yet been studied due to their low frequency [4]. In a study by Witt and Luck, it was shown that the MIT mutation is located in the starting codon of the gene, disrupts the synthesis of the trypsin inhibitor, and is inherited autosomal dominant with high penetrance [18, P. 2716–2717.].

O. Kiraly et al. [Kiraly O., Boulling A., Witt H. [et al]. Signal peptide variants that impair secretion of pancreatic secretory trypsin inhibitor (SPINK1) cause autosomal dominant hereditary pancreatitis //Hum. Mutat. 2007. Vol. 28, No. 5. P. 469–476.] New mutations were found in the first exon of the SPINK1 gene that damage the secretory signaling peptide: c. 41T4G (p.L14R) and c.36G4C (p.L12F). Both mutations were found in families with autosomal dominant inheritance of pancreatitis. Mutation p.L14R initiates rapid intracellular degradation of the trypsin inhibitor, reducing its secretion. The authors attributed these mutations not to the modifiers of the disease, but to its immediate cause.

However, the genetic status is of great importance: for example, in the presence of mutations in the genes of the pancreatic secretory trypsin inhibitor (PSTI/SPINK1) or the cystic fibrosis gene (CFTR), the risk of developing pancreatitis increases by 50 times, and if a patient with such mutations also abuses alcohol — by 200 times. The maximum risk (by a factor of 1000) was observed in a combination of genetic and environmental factors (mutation of the cationic trypsinogen PRSS1 gene and alcohol abuse or smoking) [19, 346–350.].

In Finland, only 50% of patients with the R122H mutation of the PRSS1 gene were found to have hereditary pancreatitis [Raty S., Babu M., Pelli H. et al. Human cationic trypsinogen (PRSS1) and trypsinogen inhibitor gene (SPINK1) mutation screening in a Finnish hereditary pancreatitis family. 38th Eur. Pancr. Club. 2006. 1. P34.], similar data were obtained in Europe and in the United States And vice versa, according to Keiles S., 49% of patients with pancreatitis of various etiologies have at least one mutant allele of the PRSS1, PSTI/SPINK or CFTR genes [20, 221-227.]. Among 94 patients with AP, the Italian authors found only 1 mutation (N34S) of the PSTI/SPINK1 gene and did not detect any mutation of the PRSS1 gene [Perri F., Piepoli A., Stanziale P. et al. Mutation analysis of the cystic fibrosis transmembrane conductance regulator (CFTR) gene, the cationic trypsinogen (PRSS1) gene, and the serine protease inhibitor, Kazal type 1 (SPINK1) gene in patients with alcoholic chronic pancreatitis // Eur. J. Hum. Genet. 2003. 11. 687–692.]. Considering that among the four patients with pancreatitis with genetic mutations, not a single person abuses alcohol [21, 42-47], the authors believe that the genetic basis of increased sensitivity to alcohol or the modulation of the inflammatory response of the pancreatic tissues in response to its effects remains unknown.

The cystic fibrosis transmembrane regulator (CFTR) gene is the third most common among genes with mutations associated with the development of pancreatitis [22, P. 467-474.]. CFTR is a cyclic adenosine monophosphate-sensitive anionic channel in the apical membrane of epithelial cells of some organs, including the pancreatic ducts, that controls the transport of chlorine and bicarbonates. Many of its mutations are known [23, P. 1133-1137.]. Some of them completely disrupt the functioning of the protein and cause severe clinical manifestations, others only reduce

its function [24, 134-138.]. The most severe CFTR mutations are found in cystic fibrosis, a hereditary autosomal recessive disease characterized by damage to the endocrine glands [25,1229–1256.].

When the CFTR is damaged, the transport of bicarbonates into the lumen of the duct is most disturbed, which leads to a decrease in the hydrogen index of pancreatic juice, and as a result, causes a violation of the solubilization of proteins and the transport of zymogenic granules. Equally important, acidification of the environment also contributes to the auto-activation of trypsinogen and disrupts the inactivation of trypsin [26, 134-138.]. Nevertheless, clinical manifestations of acute pancreatitis are found only in 1-2% of patients with cystic fibrosis [27, 86-95.]. Data from some studies have suggested that some mutations in the CFTR gene can play a role in pancreatic lesions and without the development of cystic fibrosis [28, 178-181]. Persons with less severe mutations that do not cause cystic fibrosis, in which the secretory function of the organ is preserved, are susceptible to pancreatitis [29, P. 951-952.]. This is attributed to the fact that most of the pancreatic tissue that supports the inflammatory process is preserved in such people [30, P. 609-620.]. In their study, J. Okcenga et al. [Ockenga J., Stuhmann M., Ballmann M. [et al]. Mutations of the cystic fibrosis gene, but not cationic trypsinogen gene, are associated with recurrent or chronic idiopathic pancreatitis // *Am. J. Gastroenterol.* 2000. Vol. 95, No. 8. P. 2061-2067.] showed that CFTR mutations are associated with chronic and acute recurrent pancreatitis. Among patients with idiopathic chronic pancreatitis, such mutations were found in 45 %, and among patients with repeated attacks of acute pancreatitis – in 38 % of cases [31, P. 372–381].

Chymotrypsin C is an enzyme of the pancreas that specifically cleaves the Leu81–Glu82 peptide bond in the cationic trypsinogen molecule, thus performing its degradation. This provides a second line of protection for organ tissue from prematurely activated trypsinogen after the trypsin inhibitor SPINK1. Since the corresponding cleavage sites are present in the molecules of anionic trypsinogen and mesotrypsin, chymotrypsin C probably provides their degradation, although there is no experimental confirmation of this yet [32, 1238-1246.]. It is suggested that CTRC mutations that reduce the activity of the enzyme or disrupt its synthesis may lead to the development of pancreatitis as a result of reduced degradation of excess trypsin [33, P. 78–82.].

Two CTRC mutations-microdeletions p. K247_R254del and p.R254W, located in exon 7, are the most common polymorphic variants found in 3.3% of patients with idiopathic chronic pancreatitis in the European population. They were also detected in 2.9 % of people suffering from alcoholic pancreatitis, which exceeded the indicator for patients with alcoholic liver disease without damage to the pancreas, which is 0.7 %. These data indicate the role of CTRC mutations in the formation of predisposition to alcoholic pancreatitis.

Chymotrypsin C (CTRC) variants that diminish activity or secretion are associated with chronic pancreatitis // *Nat. Genet.* 2008. Vol. 40, No. 1. P. 78-82.] also demonstrated that tropical pancreatitis can be caused by CTRC polymorphism. The authors identified its mutations in 14.1 % of patients with the tropical form of

chronic pancreatitis and only in 1.2 % of healthy individuals in India. The previously described mutations p. K247_R254del and p.R254W were not detected. Some authors have diagnosed tropical pancreatitis in carriers of the c.180C>T mutation, also seen in the French population [34, 889-894.], others noted the p.V235I mutation as the most frequent variant of polymorphism among patients from India (4.9 % of the examined) [35, 1602–1606.].

The calcium - sensing receptor (CASR) plays a key role in calcium homeostasis and is expressed in many tissues involved in its metabolism, including the cells of the pancreatic acinuses and ducts [36, 705-711.]. The gene encoding the calcium-sensitive receptor is located in the long arm of the third chromosome. To date, more than 70 mutations of this gene have been described, including heterozygous inactivating mutations that cause familial hypocalciuric hypercalcemia (SGH), which is considered a benign disease accompanied by an increase in the level of calcium in the blood plasma. The data that SSG is often accompanied by recurrent pancreatitis suggested a common genetic cause of the two pathologies [37, 675–680.].

P. Felderbauer et al. [Felderbauer P., Hoffmann P., Einwachter H. [et al]. A novel mutation of the calcium sensing receptor gene is associated with chronic pancreatitis in a family with heterozygous SPINK1 mutations // *BMC Gastroenterol.* 2003. Vol. 3. P. 34.] studied a family with cases of SGH and chronic pancreatitis. In patients with SGH, a sporadic mutation L173P (518T>C) in the CASR gene was found, causing the replacement of leucine with proline in the extracellular domain of the protein. The same patients suffered from recurrent attacks of acute pancreatitis, which allowed the authors to suggest the presence of a predisposition to chronic pancreatitis in individuals with this mutation. They explain the possibility of such a relationship by the fact that hypercalcemia causes premature activation of trypsinogen, which serves as a trigger factor for pancreatitis. This is confirmed by data on the increased incidence of pancreatitis in individuals with primary hyperparathyroidism (1-19 %), accompanied by an increase in the level of calcium in the blood due to the homozygous CASR mutation. However, P. Felderbauer et al. [Felderbauer P., Hoffmann P., Einwachter H. [et al]. A novel mutation of the calcium sensing receptor gene is associated with chronic pancreatitis in a family with heterozygous SPINK1 mutations // *BMC Gastroenterol.* 2003. Vol. 3. P. 34.] found that neither parathyroidectomy nor treatment with bisphosphonates led to the expected improvement in the patients' condition.

Due to the fact that proinflammatory and regulatory cytokines play an important role in the pathogenesis of acute pancreatitis, many studies have been conducted in recent years to study the role of polymorphism of genes encoding this group of proteins.

So, the researchers' attention was attracted by the CD14 protein. It exists in two forms: membrane, present on the surface of monocytes, macrophages, neutrophils, and soluble, circulating in the bloodstream. CD14 recognizes lipopolysaccharide molecules and plays an important role in immune responses by stimulating the cytokine-induced response of cells to the appearance of

lipopolysaccharide [38, 1312–1322]. Two most common polymorphisms of the CD14 gene were identified, differing in position: -260 and -651. A. Masamune et al. [39, P. 225-233.], assessing the relationship between polymorphic variants of the CD14 promoter region and pancreatic diseases in the Japanese population, found that the frequencies of the genotype –260C/T and –651C / T did not differ between healthy and patients with acute or chronic pancreatitis. Similarly, S. H. Rahman et al. did not reveal the predominance of any polymorphic variant of CD14 in patients with acute pancreatitis.

The effect of the CD14 genotype on the risk of alcohol-induced pancreatitis was also studied. Y. Chao et al. [40, P. 6043-6048.] registered a higher frequency of alleles –260C among patients with alcoholic acute pancreatitis than among healthy or suffering patients. pancreatitis of a different etiology. Another group of researchers, on the contrary, excluded the influence of CD14 polymorphism on the likelihood of developing alcoholic or biliary acute pancreatitis [41, P. 56–61.].

In addition to the CD14 protein, among the cytokines that may predispose to the development of pancreatitis, polymorphic variants of tumor necrosis factor- α continue to be studied. This protein plays the role of the main mediator of the immune response to the appearance of endotoxins. Two major polymorphic variants of the tumor necrosis factor- α gene occur when guanine is replaced with adenosine at positions -308 and -238 of the promoter region.

Despite the assumptions, these mutations occur with the same frequency among healthy and patients with acute pancreatitis [42, P. 229-234.], showed that the –308A/G mutation of this gene does not increase the risk of acute pancreatitis (similar results were obtained for the –238G/A variant).

Many polymorphic variants have been found for interleukins. The –174C/C genotype of interleukin-6 has been shown to be associated with acute biliary pancreatitis [43, P. 295-301.]. Genotype-251A/T of interleukin-8, according to some data, is more common in patients with pancreatitis, and according to others-is present with the same frequency in both patients and healthy. No differences in the frequency of different variants of the interleukin-10 genotype (–1082A/G, –819T/C, and –592A/C) and interleukin-6 (–174G/C) were found [44, P. 542–548.].

Polymorphism of genes of other cytokines, such as transforming growth factor- β 1 and gamma-interferon, according to various authors, does not play a role in the development of chronic pancreatitis, as well as alcohol-induced pancreatitis. The latter is also indicated for tumor necrosis factor- α and interleukin-10 [45, P. 162-171.]. There is evidence of the role of the heat shock protein HSP70-2 gene polymorphism in the development of pancreatitis. Thus, the mutant G-allele HSP70-2 is found with much greater frequency among patients with acute and chronic pancreatitis [46, P. 414-419.]. However, polymorphic variants of this gene are not involved in the appearance of alcoholic chronic pancreatitis [47, P. 1721–1727.].

The above indicates an undeniable relationship between certain gene mutations and certain forms of acute and chronic pancreatitis. However, many research data are

contradictory, and studies on the combinations of mutations in destructive-inflammatory diseases of the pancreas are very few.

References:

1. Pugaev A.V. Acute pancreatitis. Moscow: Profil, 2007. 335 p.].
2. Galperin E. I. Diagnostics and surgical treatment of pancreatic necrosis / / Surgery. 2003. No. 3. pp. 55-59.; Ermolov A. S. The main causes of lethality in acute pancreatitis in Moscow hospitals // Proceedings of the N. V. Sklifosovsky Research Institute of Emergency Medicine. 2001. T. 153. S. 4-14.].
3. Beger H. G., Rau B., Isenmann R. Natural history of necrotizing pancreatitis // Pancreatology. 2003. Vol. 3, No. 2. P. 93–101.].
4. Ursov S. V. Optimization of diagnostics and treatment of pancreatic necrosis / / Congress of Moscow surgeons: emergency and specialized surgical care: abstracts of reports. M., 2005. pp. 117-118., Uhl W., Warshaw A., Imrie C. [et al.]. IAP Guidelines for the Surgical Management of Acute Pancreatitis // Pancreatology. 2002. Vol. 2, No. 6. P. 565–573].
5. Savelyev V. S., Filimonov M. I., Gelfand B. R., Burnevich S. Z. Destructive pancreatitis: an algorithm for diagnosis and treatment (project). 2001. Vol. 3, no. 6. pp. 373-379.].
6. Vinnik Yu. s., Holman M. I., Popov V. O. Acute pancreatitis: pathogenesis, clinic, treatment. - Krasnoyarsk-Zelenogorsk, 1997. - 208 p., Kubyshkin V. A. Pancreonecrosis: Dis.... doctor of medical sciences. - M., 1986 – - 384 p., Sotnikov A. A. Localization of foci of hemorrhagic necrosis with different variants of the ductal system of the gland // Questions of reconstructive plastic surgery. - 2002. - No. 2. - pp. 45-49., Howes N., V. Greenhalf, S. Rutherford, et al. A new polymorphism of the R122H mutation in hereditary pancreatitis / / Kishechnik. - 2001. - Vol. 48. No. 2. - pp. 247-250.
7. Comfort M. V., Steinberg A. G. Pedigree of a family with hereditary chronic recurrent pancreatitis // Gastroenterology. 1952. Vol. 21, issue 1. pp. 54-63., Howes N., V. Greenhalf, Rutherford S. [et al.]. A new polymorphism of the R122H mutation in hereditary pancreatitis. 2001. Vol. 48, vol. 2. P. 247-250.].
8. Le Bodic L., Bignon J. D., Ragueneas O. [et al.]. The hereditary pancreatitis gene is mapped to the long arm of chromosome 7 // Hum. Mole. My wife. 1996. Vol. 5, vol. 4. Pp. 549-554., Le L. S., Schnee M., Mini-T. [et al.]. Exclusive genealogy of hereditary chronic pancreatitis // Dig. Dis. Candidate of Technical Sciences 1996. Vol. 41, vol. 7. P. 1504-1510.]
9. Baranov B.C. Genetic basis of predisposition to some frequent multifactorial diseases // Medical genetics. - 2004. - No. 3. - p. 102-112., Vinnik Yu. S., Cherdantsev D. V., Markova E. V., etc. Genetic aspects of pancreatitis // Siberian Medical Journal (Irkutsk). - 2004. - No. 2. - p. 12-17., Mayev I. V. Hereditary pancreatitis // Russian Journal of gastroenterol., hepatol. and coloproctol. - 2004. - No. 1. - p. 20-25.].
10. Markova E. V., Zotova N. V., Titova N. M., et al . Inheritance of pancreatitis: modern aspects //Actual problems of biology, medicine, and ecology. -

2004. - No. 1-3. - pp. 49-51., Howes N., Greenhalf W., Rutherford S. et al. A new polymorphism of the R122H mutation in hereditary pancreatitis // *Bowel*. - 2001. - Vol. 48. No. 2. - P. 247-250.].

11. Bernardino A. L. F., Guarita D. R., Carlos B. M., et al. Mutations of CFTR, PRSS1 and SPINK1 in the development of pancreatitis in Brazilian patients // *J. Pancreas*. - 2003. - Vol. 4. No. 5. - pp. 169-177.].

12. Chen J. M., Mercier B., Ferec C. Strong evidence that the N21I substitution in the cationic trypsinogen gene causes disease in hereditary pancreatitis // *Gut*. 1999. Vol. 45, No. 6. P. 916., Howes N., Greenhalf W., Rutherford S. [et al]. A new polymorphism for the R122H mutation in hereditary pancreatitis // *Gut*. 2001. Vol. 48, No. 2. P. 247–250., Le Marechal C., Bretagne J.F., Ragueneas O. [et al].

13. Rebours V., Levy P., Ruzsniwski P. An overview of hereditary pancreatitis // *Dig. Liver Dis*. 2012. Vol. 44, No. 1. P. 8–15..

14. Hirota M., Ohmuraya M., Baba H. Genetic background of pancreatitis // *Postgrad. Med. J*. 2006. Vol. 82, No. 974. P. 775–778.

15. Mayev I. V., Kucheryavy Yu. A. Diseases of the pancreas. Vol. 2. M.: Medicine, 2008. 560 p., Lerch M. M., Mayerle J., Aghdassi A. A. [et al]. Advances in the etiology of chronic pancreatitis // *Dig. Dis*. 2010. Vol. 28, No. 2. P. 324–329.

16. Truninger K., Witt H., Kock J. [et al.]. Mutations of the serine protease inhibitor, Kazal type 1 gene, in patients with idiopathic chronic pancreatitis // *Am. J. Gastroenterol*. 2002. Vol. 97, No. 5. P. 1133-1137.

17. Drenth J.P., de Morsche R., Jansen J.B. Mutations in serine protease inhibitor Kazal type 1 are strongly associated with chronic pancreatitis // *Gut*. 2002. Vol. 50, No. 5. P. 687–692., Witt H., Luck W., Becker M. [et al.]. Mutation in the SPINK1 trypsin inhibitor gene, alcohol use, and chronic pancreatitis // *JAMA*. 2001. Vol. 285, No. 21. P. 2716–2717.

18. Witt H., Luck W., Becker M. [et al.]. Mutation in the SPINK1 trypsin inhibitor gene, alcohol use, and chronic pancreatitis // *JAMA*. 2001. Vol. 285, No. 21. P. 2716–2717.

19. Keim V. Identification of patients with genetic risk factors of pancreatitis: impact on treatment and cancer prevention // *Dig. Dis*. 2003. 21. 346–350.

20. Keiles S, Kammesheidt A. Identification of CFTR, PRSS1, and SPINK1 mutations in 381 patients with pancreatitis // *Pancreas*. 2006. 33. (3). 221-227.

21. Irina GRIGORIEVA, Tatyana NIKITENKO, Alla YAMLIKHANOVA, Vladimir MAKSIMOV, Tatyana MIRONENKO, Mikhail VOEVODA, Alcoholic pancreatitis: gender, age, genetic features.- *Bulletin of the Russian Academy of Medical Sciences*, No. 3 (137), 2009-p. 42-47

22. LaRusch J., Whitcomb D. C. Genetics of pancreatitis // *Curr. Opin. Gastroenterol*. 2011. Vol. 27, No. 5. P. 467-474

23. Truninger K., Witt H., Kock J. [et al.]. Mutations of the serine protease inhibitor, Kazal type 1 gene, in patients with idiopathic chronic pancreatitis // *Am. J. Gastroenterol*. 2002. Vol. 97, No. 5. P. 1133-1137.

24. Kornienko E. A., Yagupova A. A. Modern ideas about the etiology of chronic pancreatitis and correction of functional pancreatic insufficiency glands // Questions of modern pediatrics. 2012. Vol. 11, no. 4. pp. 134-138.
25. Davis P. B., Drumm M., Konstantin M. W. Cystic fibrosis // Am. J. Resp. Crit. Care Med. 1996. Vol. 154, No. 5. P. 1229–1256.
26. Kornienko E. A., Yagupova A. A. Modern ideas about the etiology of chronic pancreatitis and the correction of functional insufficiency of the pancreas // Questions of modern pediatrics. 2012. Vol. 11, no. 4. pp. 134-138.
27. Shwachman H., Lebenthal E., Khaw K. T. Recurrent acute pancreatitis in patients with cystic fibrosis with normal pancreatic enzymes // Pediatrics. 1975. Vol. 55, No. 1. P. 86-95.
28. Bank S., Marks I. N., Novis B. Sweat electrolytes in chronic pancreatitis // Am. J. Dig. Dis. 1978. Vol. 23, No. 2. P. 178-181., Longnecker D. S. Pathology and pathogenesis of diseases of the pancreas // Am. J. Pathol. 1982. Vol. 107, No. 1. P. 99-121.
29. Del Rosario J. F., Putnam P. E., Orenstein D. M. Chronic pancreatitis in a patient with cystic fibrosis and clinical pancreatic insufficiency // J. Pediatr. 1995. Vol. 126, No. 6. P. 951-952.
30. Durie P. R. Pancreatic aspects of cystic fibrosis and other inherited causes of pancreatic dysfunction // Med. Clin. North Am. 2000. Vol. 84, No. 3. P. 609-620.
31. Bishop M.D., Freedman S.D., Zielenski J. [et al.]. The cystic fibrosis transmembrane conductance regulator gene and ion channel function in patients with idiopathic pancreatitis // Hum. Genet. 2005. Vol. 118, No. 3–4. P. 372–381., Ockenga J., Stuhmann M., Ballmann M. [et al.]. Mutations of the cystic fibrosis gene, but not cationic trypsinogen gene, are associated with recurrent or chronic idiopathic pancreatitis // Am. J. Gastroenterol. 2000. Vol. 95, No. 8. P. 2061–2067.
32. Zhou J., Sahin-Toth M. Chymotrypsin C mutations in chronic pancreatitis // J. Gastroenterol. Hepatol. 2011. Vol. 26, No. 8. P. 1238-1246.
33. Rosendahl J., Witt H., Szmola R. [et al.]. Chymotrypsin C (CTRC) variants that diminish activity or secretion are associated with chronic pancreatitis // Nat. Genet. 2008. Vol. 40, No. 1. P. 78–82.
34. Derikx M. H., Szmola R., de Morsche R. H. [et al.]. Tropical calcific pancreatitis and its association with CTRC and SPINK1 (p.N34S) variants // Eur. J. Gastroenterol. Hepatol. 2009. Vol. 21, No. 8. P. 889-894.
35. Paliwal S., Bhaskar S., Mani K. R. [et al.]. Comprehensive screening of chymotrypsin C (CTRC) gene in tropical calcific pancreatitis identifies novel variants // Gut. – 2013. Vol. 62, No. 11. P. 1602–1606.
36. Racz G. Z., Kittel A., Riccardi D. [et al.]. Extracellular calcium sensing receptor in human pancreatic cells // Gut. 2002. Vol. 51, No. 5. P. 705-711.
37. Pearce S. H., Wooding C., Davies M. [et al.]. Calcium-sensing receptor mutations in familial hypocalciuric hypercalcaemia with recurrent pancreatitis // Clin. Endocrinol. (Oxf). 1996. Vol. 45, No. 6. P. 675–680.

38. Rahman S. H., Ibrahim K., Larvin M. [et al.]. Association of antioxidant enzyme gene polymorphisms and glutathione status with severe acute pancreatitis // *Gastroenterology*. 2004. Vol. 126, No. 5. P. 1312–1322, Rahman S.H., Menon K.V., Holmfield J.H. [et al.].

39. Masamune A., Kume K., Kikuta K. [et al.]. –651C/T promoter polymorphism in the CD14 gene is associated with severity of acute pancreatitis in Japan // *J. Gastroenterol*. 2010. Vol. 45, No. 2. P. 225-233.

40. Chao Y. C., Chu H. C., Chang W. K. [et al.]. CD14 promoter polymorphism in Chinese alcoholic patients with cirrhosis of liver and acute pancreatitis // *World J. Gastroenterol*. 2005. Vol. 11, No. 38. P. 6043-6048.

41. Tukiainen E., Kylanpaa M. L., Puolakkainen P. [et al.]. Polymorphisms of the TNF, CD14, and HSPA1B genes in patients with acute alcohol-induced pancreatitis // *Pancreas*. 2008. Vol. 37, No. 1. P. 56–61.

42. Özhan G., Yanar H. T., Ertekin C., Alpertunga B. Polymorphisms in tumour necrosis factor alpha (TNFalpha) gene in patients with acute pancreatitis // *Mediators Inflamm*. 2010. doi: 10.1155/2010/482950.]. A meta-analysis conducted by Z. Yang et al. [Yang Z., Qi X., Wu Q. [et al.]. Lack of association between TNF-alpha gene promoter polymorphisms and pancreatitis: a meta-analysis // *Gene*. 2012. Vol. 503, No. 2. P. 229-234.

43. De Madaria E., Martinez J., Sempere L. [et al.]. Cytokine genotypes in acute pancreatitis: association with etiology, severity, and cytokine levels in blood // *Pancreas*. 2008. Vol. 37, No. 3. P. 295-301.

44. Chen J. M., Mercier B., Ferec C. Strong evidence that the N21I substitution in the cationic trypsinogen gene causes disease in hereditary pancreatitis // *Gut*. 1999. Vol. 45, No. 6. P. 916., Hofner P., Balog A., Gyulai Z. [et al.]. Polymorphism in the IL-8 gene, but not in the TLR4 gene, increases the severity of acute pancreatitis // *Pancreatology*. 2006. Vol. 6, No. 6. P. 542–548.

45. Schneider A., Barmada M. M., Slivka A. [et al.]. Analysis of tumor necrosis factor-alpha, transforming growth factor-beta 1, interleukin-10, and interferon-gamma polymorphisms in patients with alcoholic chronic pancreatitis // *Alcohol*. 2004. Vol. 32, No. 1. P. 19–24., Schneider A., Larusch J., Sun X. [et al.].

46. Srivastava P., Shafiq N., Bhasin D. K. [et al.]. Differential expression of heat shock protein (HSP) 70-2 gene polymorphism in benign and malignant pancreatic disorders and its relationship with disease severity and complications // *JOP*. 2012. Vol. 13, No. 4. P. 414-419.

47. Lee S. H., Ryu J. K., Jeong J. B. [et al.]. Polymorphisms of the MCP-1 and HSP70-2 genes in Korean patients with alcoholic chronic pancreatitis // *Dig. Dis. Sci*. 2008. Vol. 53, No. 6. P. 1721–1727.